ANTIOXIDANTS & REDOX SIGNALING Volume 17, Number 11, 2012

Mary Ann Liebert, Inc.

DOI: 10.1089/ars.2012.4702

Redox Albuminomics: Oxidized Albumin in Human Diseases

Graziano Colombo, Marco Clerici, Daniela Giustarini, Ranieri Rossi, Aldo Milzani, and Isabella Dalle-Donne

Abstract

Significance: Albumin is the major contributor to colloid oncotic pressure and also serves as an important carrier protein of many endogenous and exogenous molecules throughout the body. In blood and extravascular fluids, albumin is susceptible to different oxidative modifications, especially thiol oxidation and carbonylation. Because of its metal-binding properties and the redox properties of its Cys34 thiol, albumin displays an important antioxidant activity. As albumin is the predominant protein in most body fluids, its Cys34 represents the largest fraction of free thiols within body fluids. Recent Advances: Evidence that albumin oxidation takes place in vivo has been reported only recently. Different redox proteomic, mass spectrometric, and chromatographic techniques have shown albumin redox modifications in various human pathophysiological conditions. As a whole, most data here presented demonstrate that massive albumin oxidation occurs in vivo in different biological fluids and, to some extent, that this process is correlated to organ dysfunction. Critical Issues: Recent reports suggest that the albumin redox state may serve as a global biomarker for the redox state in the body in various human diseases. However, further study is required to elucidate the exact relationship between albumin oxidation and pathology. In addition, it is unknown if some albumin oxidized forms may also have diagnostic uses. Future Directions: Application of specific redox proteomics techniques for the characterization of oxidized albumin forms in screening studies is required. A further challenge will be to analyze how these oxidative albumin modifications are related to real impact to the body. Antioxid. Redox Signal. 17, 1515–1527.

Introduction

OXIDATIVE STRESS has been related increasingly to the onset and/or progression of a growing number of human diseases, and there is a considerable research interest in oxidative stress biomarkers for earlier prediction of disease onset and monitoring of disease progression. Increased oxidative damage has been reported for most diseases, ranging from acute to chronic ones, although the finding of false positives in this field is a quite frequent occurrence (23). A clear correlation between disease and oxidative stress is indeed far from being proven for many pathological conditions. Therefore, the identification of reliable biomarkers of oxidative damage would be of great help in basic and clinical research.

Biomarker is a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or responses to a therapeutic intervention. A few biomarkers of oxidative stress/damage are validated (16, 18, 23, 43). Proteins are major targets of oxidants and can undergo reversible redox reactions, which can also be part of their normal function, or irreversible oxidative damage at various amino acids, which are generally associated with a (permanent) loss of function (14–18, 23). Whenever ascertained, oxidative modifications may allow to establish a putative relationship between structural/functional protein alterations and pathological conditions.

Research toward a better understanding of the molecular mechanisms of a disease together with identification of specific targets of oxidative stress/damage is urgently necessary. Although tissue samples represent the best choice for understanding direct target-organ effects, there is an overwhelming, impelling need to validate more specific oxidative damage biomarkers that can be easily measured in human biological fluids obtained from noninvasive approaches, to overcome limitations of invasive monitoring (7, 43). The most accessible biological fluids for potential biomarker discovery are intravascular fluids (blood and plasma) and extravascular fluids, such as cerebrospinal fluid (CSF), synovial fluid (SF), and bronchoalveolar lavage fluid (BALF). In particular, blood

Departments of ¹Life Sciences and ²Biomedical Sciences for Health, University of Milan, Milan, Italy.

plasma is a rich source of information, because it contains a large majority of all human proteins, including albumin. Since alterations in some blood parameters can reflect alterations even in hardly achievable compartments of the body, the redox state of albumin has been analyzed in some human diseases related to oxidative stress.

Metabolism and Distribution of Human Albumin

In healthy adults, albumin synthesis occurs almost exclusively in hepatocytes ($\sim 0.2 \,\mathrm{g/kg}$ body weight per day) and accounts for 10% of total liver protein synthesis. Whether human serum albumin (HSA) found in milk is produced by the mammary gland or derived from the circulation has not yet been resolved (19, 56). Recently, convincing evidence of albumin synthesis in human microglial cells has been provided (19). There is no storage of newly synthesized albumin, only a small amount (<2%) in transit during its 30-min trip through hepatocytes. Under physiological conditions, only 20%–30% of hepatocytes produce HSA and its synthesis can therefore be increased on demand by a factor of 200%-300% (56). The colloid osmotic pressure and the osmolality of the interstitial liquid around the hepatocytes is the most important regulator of HSA biosynthesis, which is also regulated by the nutritional status and hormones, including insulin, glucagon, cortisol, and thyroid hormones (19, 56). Under physiological conditions, a steady state exists between albumin synthesis and metabolism. The amount of albumin metabolized daily is proportional to the plasma concentration: a fixed percentage of $\sim 10\%$ of HSA content is metabolized per day. The catabolism of HSA takes place in several organs, primarily in muscle, skin, and the liver, after \sim 20 days from its synthesis at a rate of about 14 g per day in a 70 kg healthy adult, or 4% of whole-body protein turnover (19, 56).

Albumin distribution is adequately described by a twocompartment model where about 40% is taken up by the intravascular and 60% by the extravascular space (Fig. 1). Intravascular (blood plasma) HSA is the most abundant protein (mean concentration $\sim 43 \,\mathrm{mg/ml}$, range $35-52 \,\mathrm{mg/ml}$, i.e., \sim 0.6 mM), accounting for 50%–60% of total plasma proteins in healthy humans (56). Measurement of albumin concentration in the extravascular fluids has been performed in healthy humans (Supplementary Table S1; Supplementary Data are available online at www.liebertpub.com/ars). The total exchange rate between intravascular and extravascular fluid (transcapillary escape rate, i.e., the movement of HSA across the capillary wall) amounts to \sim 5% of the intravascular albumin content per hour (Fig. 1); HSA returns to the bloodstream at an equivalent rate through the lymphatic system via the thoracic duct, with a circulation half-life of $\sim 16 \, h$ (19, 56).

Structure and Functions of Human Albumin

HSA is a single polypeptide of 585 amino acids (66.5-kDa), including 35 cysteine residues forming 17 intramolecular disulfide bonds, with the only free sulfhydryl group located at Cys34 (56). Reduced form of HSA (Alb-SH) is also known as human mercaptalbumin (HMA, with Cys34 containing a free thiol). Within the tertiary protein structure of HSA, Cys34 resides in a unique microenvironment close to three ionizable

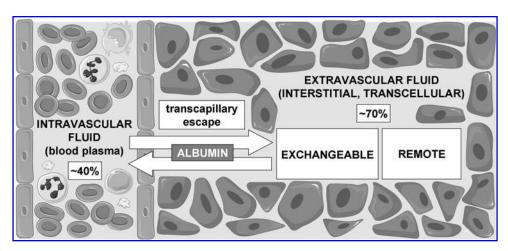


FIG. 1. Albumin distribution in human extracellular fluids. Body fluids are divided between intracellular fluid ($\sim 2/3$ of body water) and extracellular fluid (\sim 1/3 of body water). Extracellular fluid is divided between intravascular fluid (or blood plasma, ~1/5 of extracellular fluid) and extravascular fluid. Extravascular fluid can be further partitioned between interstitial fluid ($\sim 4/5$ of extracellular fluid) and transcellular fluid. Interstitial fluid (or tissue, or intercellular fluid) consists of all the bits of fluid that lie in the interstices of all body tissues. This is also a virtual fluid, that is, it exists in many separate small bits, but it is considered as a pool of fluids of uniform composition in the one location. The interstitial fluid dampens all the cells in the body and is the link between the intracellular fluid and the intravascular one. Transcellular fluid is a small compartment that represents all those body fluids formed from the transport activities of cells. It is contained within epithelial-lined spaces and includes fluids such as CSF, synovial fluid, bladder urine, and ocular fluid (aqueous humor of the eye). The plasma compartment holds about 40% of the total body albumin pool, the rest being in extravascular compartments. A portion of extravascular albumin is tissue-bound (remote component) and is therefore unavailable to the circulation. Albumin cyclically leaves the circulation, passing through the endothelial barrier at the level of the capillaries, and translocating into the interstitial fluid and returns to the bloodstream through the lymph system via the thoracic duct. The circulation half-time for this process takes ~16h. The movement of HSA across the capillary wall is defined as the transcapillary escape (at the rate of ~5% per hour), meaning that 5% of intravascular HSA hourly leaves the intravascular compartment (56). CSF, cerebrospinal fluid; HSA, human serum albumin.

residues, Asp38, His39, and Tyr84 (69), which affect the ionization state of Cys34, thus modulating its reactivity (19, 55). As a result, Cys34 has an unusually low p K_a (<6.7 $vs. \sim 8.0$ – 8.5 for thiols in most other proteins) and exists primarily in the highly nucleophilic thiolate form (72). However, HSA presents pH-dependent structural transitions that impact on thiol reactivity, and consequently, the Cys34 p K_a value remains controversial (77). Moreover, the reactivity of the Cys34 thiol is affected by the presence of ligands. For example, fatty acids, which bind to sites relatively distant from Cys34, induce conformational changes at the thiol site and increase the thiol reactivity toward disulfides (77). Indeed, the three-dimensional (3D) structure of lipidated albumin shows increased exposure of Cys34 compared to fatty acid-free albumin (19). The enhanced reactivity of the albumin Cys34 thiol and its sulfenic derivative at neutral pH upon fatty acid binding can be explained by considering that the corresponding conformational changes that increase thiol exposure increase both the availability of the thiolate due to a lower apparent p K_a and also loosen steric constraints for reactions (77). Considering that situations that tend to increase the amount of fatty acids bound to HSA (such as diabetes mellitus) are associated with oxidative stress, a better antioxidant ability of HSA could constitute a protective adaptation. Indeed, oxidized forms of HSA are found increased under these situations (Table 1). The high reactivity of HSA toward electrophilic reactive carbonyl species is due to several accessible nucleophilic residues, mainly Cys34, followed by His146 and Lys199 (3).

Albumin can also bind metal ions such as Cu(II), Pt(II), Ag(I), Hg(II), and Co(II), in up to four sites with different specificities (56), which makes it susceptible to carbonylation *via* metal-catalyzed oxidation reactions.

The secondary structure of HSA is dominated by α -helices (68%), without any β -sheet. HSA is arranged in a globular heart-shaped conformation, organized in three homologous domains (I, II, and III), each consisting of two subdomains, A and B (12, 19, 73).

Albumin is responsible for about 80% of intravascular colloid oncotic pressure and predominantly contributes to the interstitial colloid osmotic pressure; it also modulates the capillary permeability, neutrophil adhesion, and activation (19, 52). HSA excellently binds and transports both endogenous and exogenous molecules, including longchain fatty acids, cholesterol, bile pigments, hormones, proteins and peptides (*e.g.*, bilirubin, ceruloplasmin, and apoliporoteins), nitric oxide, many drugs (*e.g.*, ibuprofen

Table 1. Albumin Cys34 Redox State in Human Diseases and Pathophysiological Conditions

Condition	Thiol modification	Source	Method	References
Focal segmental glomerulosclerosis	Alb-SO ₃ H	Plasma	LC-ESI-MS/MS	(8, 43, 44)
Mild cognitive impairment, Alzheimer's disease	Alb-SSR	Plasma	HPLC with fluorescence detection ^a	(24)
Hemodialysis patients with or without diabetes mellitus	Alb-SSR	Plasma	HPLC with fluorescence detection ^a	(37 ^b , 38 ^{b,c} , 67 ^b)
Aging of healthy humans	Alb-Cys (22,62), Alb-SSR (48)	Plasma	HPLC with fluorescence detection ^{a,d}	$(22^{\rm d}, 48^{\rm a}, 62^{\rm d})$
Senile cataract	Alb-SSR, AÌb-ŚO₂H, Alb-SO₃H	Aqueous humor	HPLC with fluorescence detection ^a	(29)
Pregnancies with intrauterine growth restriction	Alb-Cys	Maternal plasma	HPLC and ESI-MS	(4)
Gestational diabetes mellitus	Alb-Cys	Amniotic fluid	HPLC and ESI-MS	(6)
Chronic kidney disease	Alb-SSR, Alb-SO₂H, Alb-SO₃H	Serum	HPLC with fluorescence detection (73) or photo diode alley detector (72)	(72, 73)
Liver cirrhosis	Alb-SO ₂ H, Alb-SO ₃ H	Serum	HPLC with fluorescence detection ^a	(21, 77)
Advanced liver disease (liver cirrhosis, acute- on-chronic liver failure)	Alb-Cys, Alb-SO ₂ H, Alb-SO ₃ H	Plasma	HPLC with fluorescence detection ^a	(49, 50)
Diabetes mellitus, proliferating diabetic retinopathy	Alb-SSR	Plasma	HPLC with fluorescence detection ^a	(52)
Primary nephrotic syndrome	Alb-SO ₃ H	Plasma	LC-ESI-MS/MS	(43)

^a280/340 nm.

^bHemodialysis patients with diabetes mellitus.

^cHemodialysis patients without diabetes mellitus.

^dof the thiol-mBrB conjugate.

Alb-Cys, S-cysteinylated albumin (S-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1]); Alb-SH, mercaptalbumin; Alb-SO₂H, albumin sulfinic acid (also called human nonmercaptalbumin2 [HNA-2]); Alb-SO₃H, albumin sulfonic acid (also called human nonmercaptalbumin2 [HNA-2]); Alb-SOH, albumin sulfenic acid; Alb-SSR, albumin-mixed disulfide (S-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1]); LC-ESI-MS/MS, liquid chromatography—electrospray ionization tandem mass spectrometry; HPLC, high performance liquid chromatography.

and warfarin), ion metals (19, 20, 52), as well as reactive carbonyl species (3, 11).

Antioxidant activity has been recognized as a significant function of albumin in the plasma, a body compartment exposed to continuous oxidative stress where only a limited amount of antioxidant enzymes are available (59). The antioxidant activity of HSA results from the redox properties of the Cys34 residue and from its metal-binding properties. Due to the high plasma concentration of HSA, its Cys34 sulfhydryl group represents the largest fraction (\sim 80%, corresponding to \sim 500 μ M) of all free thiols in plasma, thus conferring a major role in the plasma antioxidant capacity to HSA (52, 63). In addition, as albumin is the predominant protein within BALF (which is representative of components in alveolar space), CSF, and SF (37, 48, 64), it may be the major source of free thiols in BALF, CSF, and SF as well. Therefore, Alb-SH may be an important endogenous antioxidant within the body.

HSA also displays an indirect antioxidant activity by binding several compounds, including oxysterols, bilirubin, and homocysteine (52, 63).

Thiol Oxidation in Human Albumin

The albumin Cys34 thiol exists in both reduced and oxidized forms (Fig. 2). In plasma of healthy young adults, 70%–80% of total HSA contains the free sulfhydryl group of Cys34. About 20%–30% of the Cys34 forms a reversible mixed disulfide with low-molecular-weight thiols, like cysteine, homocysteine, cysteinylglycine, or glutathione, generating Sthiolated albumin (Alb-SSR, also called human nonmercaptalbumin1 [HNA-1]), the predominant modification being S-cysteinylated albumin (Alb-Cys) (5, 27, 31, 65). A small fraction of the Cys34 (2%–5%) is more highly oxidized to the sulfinic (Alb-SO₂H) or sulfonic acid (Alb-SO₃H) form (79, 80), which are also called human nonmercaptalbumin 2 (HNA-2) and can cause a loss of protein function. The sulfenic acid form (Alb-SOH) is a central intermediate in the formation of oxidized al-

bumin species and increases under pathological conditions (10, 79, 80). Alb-SOH is often included within the HNA-2 fraction. The Cys34 thiol can also become *S*-nitrosated (78).

While *in vitro* studies have long supported the antioxidant role of HSA in plasma, evidence that albumin oxidation takes place also *in vivo* has been reported only recently. In particular, the advent of potent mass spectrometric techniques applied to albumin, together with chromatographic analytical approaches, has made detection of albumin redox modifications in different pathophysiological conditions possible, where decreases in HMA correlate with increases in oxidized HSA forms (mixed disulfides, HNA-1, and higher oxidation states, HNA-2) (Table 1). These oxidized species are not present when albumin is secreted from hepatocytes, and thus they may constitute potential biomarkers of the involvement of the oxidative stress/damage in human disease and of the antioxidant activity of the albumin Cys34.

Studies on plasma HSA performed by means of liquid chromatography-electrospray ionization tandem mass spectrometry (LC-ESI-MS/MS) demonstrated for the first time in vivo massive and stable oxidation of Alb-SH into Alb-SO₃H in nephrotic syndrome and focal segmental glomerulosclerosis (8, 44, 45), a renal disease characterized by sclerosis of the glomerular tuft, for which an increased oxidative stress has been hypothesized. This oxidation produces relevant changes to the protein structure and is thought to play a role in the pathogenesis of the proteinuria associated with focal segmental glomerulosclerosis (44, 45). Thiol oxidation to -SO₃H increases the net negative charge of albumin as demonstrated by using the electrophoretic titration curve, a 2D electrophoretic technique where proteins move perpendicularly to a pH gradient generated by a stack of carrier ampholytes, generating sigmoidal pH/mobility curves proportional to their chemical ionization (8, 44).

Oxidative modifications of HSA Cys34 have been reported in advanced liver disease. Marked increases in the percentages of HNA-1 and/or HNA-2, and a corresponding decrease

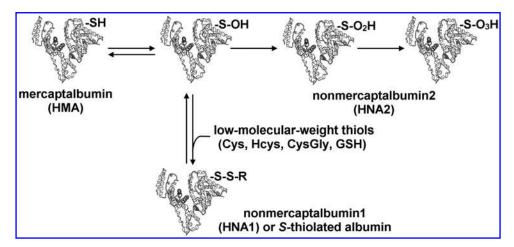


FIG. 2. Main oxidative modifications of the albumin Cys34 thiol. Oxidation of albumin Cys34 can result in the formation of a sulfenic (Alb-SOH), sulfinic (Alb-SO₂H), or sulfonic (Alb-SO₃H) acid derivative, the former of which is a reversible intermediate. Reversible S-thiolated albumin can be derived from a reaction between Alb-SOH and low-molecular-mass thiols, beside from other reaction mechanisms (15). Ribbon representation was drawn with the Swiss-PdbViewer using the PDB-file PDB-ID 1BJ5 (12). Alb-SH, mercaptalbumin; Alb-SO₂H, albumin sulfinic acid (also called human non-mercaptalbumin2 [HNA-2]); Alb-SO₃H, albumin sulfonic acid (also called human nonmercaptalbumin2 [HNA-2]); Alb-SOR, albumin-mixed disulfide (S-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1]).

in the Alb-SH percentage, were measured by high performance liquid chromatography (HPLC) with UV detection in both acute-on-chronic liver failure (50, 51) and liver cirrhosis (21, 50, 51, 81), these differences being even more pronounced in the former (50, 51). The redox state of HSA was also used as a marker to investigate the current status of oxidative stress in patients with chronic renal failure. The fraction of oxidized HSA forms (HNA-1 and HNA-2) is elevated in correlation with the degree of renal dysfunction, even before dialysis (75, 76). In addition, albumin oxidation is a powerful independent risk factor for a serious cardiovascular disease incidence and mortality among patients on hemodialysis (75). These reports suggest that the redox state of HSA is a good marker to investigate the current status of oxidative stress in patients with chronic renal failure, which is enhanced in correlation with the level of renal dysfunction.

S-thiolated albumin increases during aging and in different kinds of diseases characterized by oxidative stress, as measured by HPLC with fluorescence detection (Table 1). The percentage of Alb-SSR significantly increases with age in healthy people (22, 49, 65), in particular that of Alb-Cys (22, 65). This indicates that the antioxidant efficiency of the protein thiol pool in plasma decreases with age, causing an increased risk of irreversible oxidation of protein sulfhydryl groups and, therefore, a depletion and/or impairment of the antioxidant capacity of plasma.

Plasma Alb-SSR is increased in patients with mild cognitive impairment and Alzheimer's disease (24). Although plasma Alb-SSR is not a specific marker for neurodegenerative disease, this biomarker of oxidative stress (together with carbonylated proteins and malondialdehyde) was proposed to be a more useful marker for neurodegenerative disorders than homocysteine, vitamin B12, or folate (24). Increased levels of Alb-SSR as a systemic marker of oxidative stress have also been measured in the plasma of hemodialysis patients (38, 70), of patients with diabetes mellitus and, to an higher extent, of those with proliferating diabetic retinopathy, which also showed marked increases in the irreversibly oxidized HSA forms (53). In addition, levels of Alb-SSR and HNA-2 were found markedly increased in the aqueous humor of patients with senile cataract with no complications (30). However, further study is required to elucidate the eventual relationship between the redox state of the aqueous humor albumin and cataract formation.

Elevated levels of Alb-Cys were detected by HPLC coupled with ESI-time-of-flight (TOF)-MS in maternal plasma of pregnancies with intrauterine growth restriction (4) and by HPLC coupled with nanoESI-quadrupole-TOF-MS in amniotic fluid (where albumin is the most abundant protein) of pregnancies associated with gestational diabetes mellitus (6).

Albumin Carbonylation

Many studies have shown the presence of elevated levels of carbonylated albumin (Alb-CO) in various human diseases and pathophysiological conditions by means of spectrophotometric and immunochemical methods (enzyme-linked immunosorbent assay [ELISA], or sodium dodecyl sulfate-polyacrylamide gel electrophoresis [SDS-PAGE] combined with Western blot (OxyBlot) as well as redox proteomics (2D polyacrylamide gel electrophoresis [2-DE], combined with Western blot [2D-OxyBlot], eventually coupled to the MS

analysis) (Table 2). Spectrophotometric and most immunochemical assays rely on the use of 2,4-dinitrophenylhydrazine (DNPH) to derivatize protein carbonyl groups, which forms a stable 2,4-dinitrophenylhydrazone (DNP) adduct that is then detected, respectively, by either absorbance at 366–370 nm or by using an anti-DNP antibody (17).

Albumin is the main plasma carbonylated protein in chronic kidney disease and hemodialysis patients, as evidenced by using for the first time a Western blotting technique to detect differential susceptibility of plasma proteins to carbonyl formation in vivo in human disease (26). Carbonylation, also associated to dityrosine formation (34), alters the structural and functional properties of uremic HSA (34, 38), suggesting that oxidation of plasma HSA might adversely affect its vasculoprotective effects in hemodialysis patients. A further study also demonstrated increased albumin modification by advanced glycation end products (AGEs), and that the amount of both Alb-CO and AGE-modified HSA was the highest in hemodialysis patients with diabetes mellitus (39), a pathology where increased levels of serum Alb-CO were further confirmed (62). Increasing albumin carbonylation parallels the severity of chronic kidney disease and is even higher in the plasma of hemodialysis patients, as detected by the OxyBlot assay, indicating that oxidative stress in such patients gradually increases during the development of the disease (40). Alb-CO was also detected by the OxyBlot assay in urine of patients with chronic kidney disease (1) as well as in the plasma and urine in diabetic nephropathy (2).

There is evidence that oxidative damage occurs in rheumatoid diseases. OxyBlot showed that levels of serum Alb-CO were significantly higher in patients with rheumatoid arthritis than in controls (32). Furthermore, by means of two-stage electrophoresis, by which serum proteins were separated by standard agarose gel electrophoresis, submitted to reaction with DNPH, and resolved in the second stage by SDS-PAGE, followed by OxyBlot, albumin and γ -globulins were found to be the most carbonylated proteins in plasma of juvenile idiopathic arthritis (84).

Progressive increase of plasma Alb-CO levels, associated with increased oxidation of Cys34 (Table 1), was found by ELISA and OxyBlot assays, with increasing severity of liver cirrhosis and acute-on-chronic liver failure (51). The increase in total serum protein carbonyl levels in hepatocellular carcinoma patients (61) as well as in lung cancer patients (60) was largely due to an increase in Alb-CO levels.

In humans, a major cause of chronic oxidative stress is cigarette smoking. By the OxyBlot analysis, the major carbonylated protein in the BALF of older smokers with long-term smoking histories was found to be albumin, even in the absence of lung diseases (46, 74). Parenchymal tissue from chronic obstructive pulmonary disease (COPD) patients, who were current smokers, contained lower levels of total albumin, but had proportionally greater levels of Alb-CO, compared to patients with the normal lung function, as determined by ELISA and OxyBlot (25). Lung tissue from current smokers was also found to contain lower levels of albumin, which was highly carbonylated compared to lung tissue from ex-smokers and nonsmokers. Decreased levels of albumin in current smokers with COPD could contribute to the excessive accumulation of oxidants, which would ultimately lead to oxidative tissue injury. Lung tissue from exsmokers had the same mean concentration of Alb-CO as

TABLE 2. CARBONYLATED ALBUMIN IN HUMAN DISEASES AND PATHOPHYSIOLOGICAL CONDITIONS

Condition	Source	Method	References
Older smokers with long-term smoking histories, even in the absence of lung diseases	BALF	OxyBlot	(45, 71)
Type 1 diabetes mellitus	Serum	Spectrophotometric assay	(59)
Current smokers with chronic obstructive pulmonary disease	Parenchymal lung tissue	ELISA and OxyBlot	(25)
Allergic airway inflammation (asthma)	Sputum	OxyBlot	(46)
Chronic kidney disease	Plasma (26,39), urine (1)	OxyBlot and ELISA, spectrophotometric assay	(1, 26, 39)
Hemodialysis patients	Plasma	OxyBlot, ELISA, spectrophotometric assay	(26, 33, 37, 38)
Advanced diabetic nephropathy	Plasma, urine	OxyBlot	(2)
Guillain–Barrè syndrome	Cerebrospinal fluid	Redox proteomics + MALDI-TOF/ TOF MS	(13)
Ischemic stroke	Cerebrospinal fluid	Redox proteomics+MALDI-TOF/ MS	(40)
Juvenile arthritis	Plasma	Spectrophotometric assay and two- stage electrophoresis ^a + OxyBlot	(78)
Lung cancer	Serum	Spectrophotometric assay	(57)
Systemic lupus erythematosus	Serum	ELISA, redox proteomics + MS	(41, 65)
Liver cirrhosis, acute-on chronic liver failure	Plasma	ELISA, OxyBlot	(50)
Hepatocellular carcinoma	Serum	Spectrophotometric assay	(58)
Rheumatoid arthritis	Serum	Spectrophotometric assay and OxyBlot	(31)
Nondiabetic hemodialysis patients	Plasma	OxyBlot+MS MALDI-TOF/ MS+nanoLC-MS/MS	(53, 54)
Chronic lung diseases in children	BALF	Redox proteomics	(68)

^aStandard agarose gel electrophoresis, after which proteins in the gel were submitted to reaction with DNPH and resolved in the second stage by SDS-PAGE.

BALF, bronchoalveolar lavage fluid; MALDI-TOF, matrix-assisted laser desorption/ionization time of flight; DNPH, 2,4-dinitrophenylhydrazine; ELISA, enzyme-linked immunosorbent assay; SDS-PAGE, sodium dodecyl sulfate–polyacrylamide gel electrophoresis.

nonsmokers (25), suggesting that smoking cessation could prevent albumin carbonylation, contributing to the restoration of the oxidant/antioxidant balance within the lung.

Oxidative stress is also involved in asthma. Carbonylation of sputum proteins in adult asthmatic patients and healthy controls was assessed by using OxyBlot (47). Albumin and α 1-antitrypsin were specifically carbonylated in asthmatics, and Alb-CO was significantly correlated with sputum eosinophilia.

Recently, albumin carbonylation in different human diseases has been clearly documented by redox proteomics. Redox proteomics analysis, in combination with MS protein identification, of serum samples from patients afflicted with the autoimmune disease systemic lupus erythematosus showed albumin and immunoglobulins to be particularly carbonylated (42). In patients with stable disease activity, higher long-term protein carbonylation correlated with higher long-term disease activity, whereas patients with variable disease activity exhibited varying correlations between protein carbonylation and disease activity markers (42). These results confirm and extend previous findings (68) and further support a role for oxidative stress in this disease pathogenesis.

CSF is produced in the brain mainly by cells in the choroid plexus at a rate of $500\,\text{ml/day}$ and contains $\sim\!0.3\%$ plasma proteins, mainly albumin and immunoglobulins. It is the only body fluid in direct contact with the brain. Thus, any changes

in the CSF composition will reflect pathological processes and make CSF a potential source of biomarkers for different diseases. A recent study used redox proteomics to analyze the extent of carbonylation and, thus, of oxidative damage of CSF proteome in patients with Guillain-Barrè syndrome, an acute and immune-mediated inflammatory disease affecting the peripheral nervous system (13), evidencing a major sensitivity to carbonylation for albumin, α-glycoprotein, and Fab fragments from an IgM globulin (Fig. 3). Brain biomarkers of oxidative stress need to be identified to better understand and treat brain damage in human stroke patients. Redox proteomics was recently used to identify carbonylated proteins in the CSF of stroke patients with acute ischemic brain injury (41). Albumin was the only protein that was markedly carbonylated in the CSF from stroke patients as compared with controls (Fig. 4). Carbonylation of albumin normalized to the total albumin from each relevant group was significantly increased in CSF from stroke patients compared with the control group, and Alb-CO was significantly correlated to clinical outcomes. This finding in humans agrees with the experimental results obtained using an animal model of ischemic stroke. These results suggest that Alb-CO in CSF can be utilized as a biomarker of oxidative stress in stroke patients and animal models, thereof (41).

OxyBlot coupled to matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF)/MS mass fingerprinting identification of carbonylated proteins recently confirmed

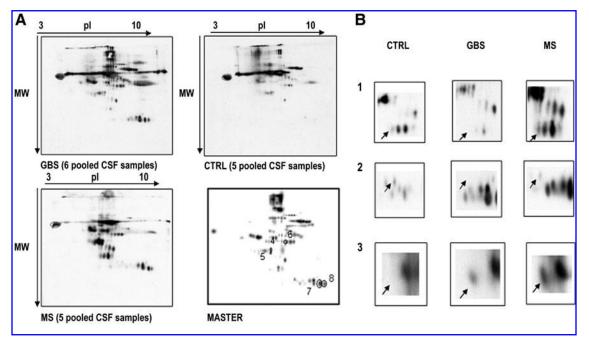


FIG. 3. Redox proteomic analysis of protein carbonylation in the CSF of patients with GBS. (A) Two-dimensional (2D) Western blotting of derivatized proteins detected with anti-DNP antibodies. Grey circles indicate internal standard. A synthetic master gel image is also presented: black circles indicate differentially carbonylated protein spots. (B) Zoomed areas of 2D Western blotting showing spots absent or present in the clinical groups (arrows). Reprinted from D'Aguanno *et al.* (13), Copyright © 2010, with permission from Elsevier. GBS, Guillain–Barrè syndrome; MW, molecular weight; CTRL, control; MS, multiple sclerosis; DNP, 2,4-dinitrophenylhydrazone.

previous observations that albumin is the major plasma protein target of oxidative/carbonyl stress in chronic hemodialysis patients (54). This redox proteomic study also first showed that albumin carbonylation in uremic blood mainly results from protein modification by carboxymethyl-lysine, in agreement with the occurrence of carbonyl stress in uremia

and hemodialysis patients. A further study demonstrated, by OxyBlot coupled to MALDI-TOF/MS mass fingerprinting and nano-LC-MS/MS analysis, that postdialysis plasma carbonylated-protein levels are significantly increased compared to predialysis levels, and that cabonylation targets almost a dozen of plasma proteins, among which albumin is the most

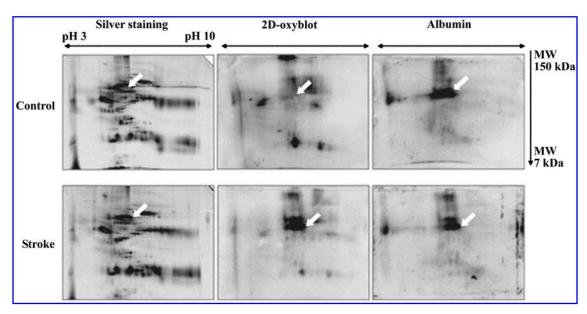


FIG. 4. 2D OxyBlot analysis of protein carbonylation in the CSF of stroke patients. 2-DE images of protein samples from CSF of control or stroke patients labeled with silver staining (*left panel*). 2D OxyBlots of protein samples immunolabeled with an anti-DNP antibody (*middle panel*). After stripping, the same membrane was immunolabeled with an anti-HSA antibody (*right panel*). Arrow indicates the albumin spot. Reprinted from Moon *et al.* (41), Copyright © 2011, with permission from John Wiley and Sons. 2-DE, two-dimensional polyacrylamide gel electrophoresis.

susceptible to carbonyl formation (55). Furthermore, the *in vitro* finding that even low concentrations of Alb-CO may have proatherogenic properties indicates a potential role for protein carbonylation in the early atherogenic events of chronic uremia.

Significantly higher levels of protein carbonylation than in healthy controls were determined in BALF of children with interstitial lung disease, gastroesophageal reflux disease, and pulmonary alveolar proteinosis, although with significant interindividual variations. Surfactant protein A, α 1-antitrypsin, and, mainly, albumin were the proteins most sensitive to carbonylation, whose extent was proportional to the count of neutrophils in BALF (71).

Albumin Nitration

Most data show that protein tyrosine nitration is a selective process where only a few proteins become nitrated, and only one or a few Tyr residues are modified within each protein. While HSA contains 18 Tyr residues, an *in vitro* study showed that only Tyr138 and Tyr411 are particularly susceptible to nitration (28).

Using a sensitive, quantitative ELISA, nitrated albumin (nitroalbumin) levels were assayed in plasma samples collected during the first hour, at day 1 and at day 4 of life from term newborns suffering from perinatal asphyxia (83). Nitroalbumin levels at day 1 were significantly increased in infants who developed moderate or severe encephalopathy compared to those who had a normal neurological evolution or developed mild encephalopathy. In contrast, the nitroalbumin concentration at day 1 was not associated with systemic complications (83). Plasma nitroalbumin at day 1 was thus specifically associated with perinatal hypoxicischemic cerebral injury. The nitroalbumin concentration increased with the severity of neonatal encephalopathy, adding support to the hypothesis that reactive nitrogen species (RNS) toxicity plays a role in hypoxic-ischemic brain injury. Therefore, plasma nitroalbumin seems to be a specific marker of neurological injury after perinatal asphyxia and is an easily measurable, noninvasive marker of nitrative stress usable as a secondary end-point in neuroprotective clinical trials. Increased plasma nitroalbumin is also associated with early transient hypoglycemia in preterm newborns, as determined by ELISA (82).

Advanced Oxidation Protein Products

Elevated levels of advanced oxidation protein products (AOPPs), which arise from the reaction between chlorinated oxidants and plasma proteins, have been described in several human diseases (Supplementary Table S2) (67). AOPPs are dityrosine-containing and cross-linking protein products formed mainly during oxidative burst of activated neutrophils via the myeloperoxidase reaction. Plasma levels of AOPPs closely correlate with levels of dityrosine and pentosidine, an AGE product closely related to oxidative stress; indeed, AOPPs contain dityrosine, protein carbonyls, and pentosidine in an aggregate-forming or monomeric form (9) (Fig. 5). Utilizing indirect techniques, albumin was identified as the main AOPP product in plasma of chronic uremic patients (9). Notwithstanding, a lack of chemical structural completeness did not allow any conclusion. In addition, recent data appear to indicate that oxidized fibrinogen is the key

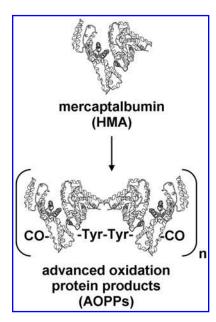


FIG. 5. Schematic representation of AOPPs. Albumin was identified as the main AOPP product in plasma of chronic uremic patients (9), although more recent data suggest that oxidized fibrinogen would be the main protein forming AOPPs in the human plasma (66, 67). AOPPs contain highly oxidized proteins, characterized by carbonylation, dityrosine cross-linking, and high molecular weight. Ribbon representation was drawn with the Swiss-PdbViewer using the PDB-file PDB-ID 1BJ5 (12). AOPPs, advanced oxidation protein products.

molecule responsible for AOPP formation in the human plasma (66, 67). However, a role of AOPPs in atherosclerotic cardiovascular events in patients with chronic renal failure and, more in general, in the inflammatory process that is associated with uremia and hemodialysis is hypothesized (36, 67). In particular, *in vivo* and *in vitro* evidence that AOPPs are proinflammatory mediators has been provided (36), and prooxidant–proinflammatory properties of AOPPs were evidenced in animal models of chronic kidney failure (33, 35, 69), suggesting a vicious cycle with amplification of oxidative stress and inflammation (Fig. 6). In the adults with chronic renal insufficiency, AOPPs accumulate proportionally with the decline in renal function and are also proposed as predictors of survival in these patients (67).

Concluding Remarks

In vivo albumin oxidation in relation to various human diseases has been proven, by direct techniques, only recently. Demonstration of oxidized HSA in biological fluids has stimulated studies aimed at defining its levels in relation to various human diseases potentially linked to oxidative/nitrative stress. High oxidized HSA levels have been assayed in vivo in pathologies where reactive oxygen species/RNS seem to play a pathogenic role, thus demonstrating the HSA antioxidant role also in vivo. Moreover, HSA purified from hemodialysis patients had decreased the free radical-scavenging ability and reduced drug-binding properties, thus confirming that oxidation-induced structural changes affect HSA- binding properties and antioxidant activity (31, 38, 52). Disturbed binding/transport properties of oxidized albumin may impair

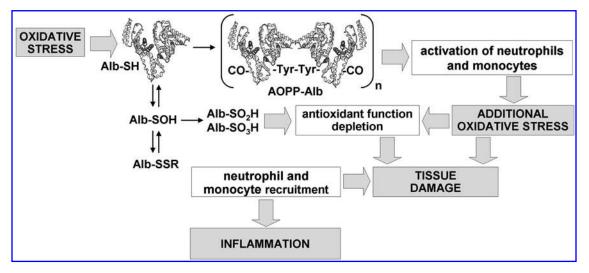


FIG. 6. Schematic representation of AOPPs as proinflammatory mediators. Pro-oxidant and/or proinflammatory properties of AOPPs were evidenced both in hemodialysis patients (36) and in animal models of chronic kidney failure (33, 35, 69), suggesting a vicious cycle with amplification of oxidative stress and inflammation. Schematic diagram showing the concomitant contribution of Alb-SO₂H, Alb-SO₃H, Alb-CO, and dityrosine-containing cross-linked AOPPs to production of additional oxidative stress and tissue damage/inflammation via activation and recruitment of neutrophils and monocytes. Ribbon representation was drawn with the Swiss-PdbViewer using the PDB-file PDB-ID 1BJ5 (12). Alb-CO, carbonylated albumin; Alb-SSR, albumin-mixed disulfide (S-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1]).

intercellular/interorgan traffic of endogenous compounds, but also delivery of drugs that are crucial in the treatment of human disease, and this may at least in part explain the altered behavior of drugs in some pathological situations.

Indeed, although the albumin Cys34 thiol does not react particularly fast with oxidants, it can still be considered an important intravascular scavenger due to its very high concentration in human plasma, particularly considering that plasma is scarce in antioxidant defenses and low molecular weight thiols. In this regard, the increase of oxidized HSA forms in pathological conditions can be considered as a proof of concept of the scavenger activity of albumin.

Approximately 10 g/h of HSA travels in and out of blood vessels (29); this means that within an hour, approximately half of intravascular HSA is replaced with extravascular HSA. As albumin is quickly distributed between blood and the extravascular compartment, it may serve as a global marker for the redox state in the body. Interestingly, nitroalbumin has been reported to be transported across the endothelial barrier two to four times faster than native albumin (58).

Current research progress on the detailed measurement of oxidative damage to proteins by different redox proteomics and MS technologies suggests that many more biomarkers are likely to be developed and clearly validated in the near future. As a whole, most data here presented demonstrate that massive albumin oxidation occurs in vivo in different biological fluids, and, to some extent, that this process is correlated to organ dysfunction in human pathology. We hope that these findings put the basis for developing/applying specific analytical techniques for the characterization of oxidized albumin forms in screening studies, also considering that in a number of studies, albumin is usually ignored, since it is generally removed from plasma/serum, CSF, or other biological tissue fluids to detect lower-abundance proteins. However, the potential use of albumin as a biomarker is presently not fully clear: is albumin oxidation only a marker of oxidative stress, or could it be reasonably considered a specific disease marker? In that light, the finding that nitroalbumin concentration is directly correlated with the severity of neurological morbidity after perinatal asphyxia indicates that nitroalbumin seems to be a reliable, early, and easily obtainable marker of long-term effects on the cognitive and behavioral consequences for the survivors of moderate-to-severe neonatal encephalopathy, beside a marker of RNS generation and nitrative stress occurrence (83). Analogously, the finding that Alb-CO was the unique carbonylated proteins found in CSF of patients with acute ischemic stroke, whose increased levels were positively correlated with neurologic indicators (41), suggests that HSA carbonylation in CSF after stroke may be able to act not only as a biomarker providing information of oxidative stress status but also as an important clue that will allow interpretation of the diagnostic outcomes of stroke.

A clear benefit of using oxidized albumin as a biomarker is that the highly abundant HSA concentration in most biological fluids allows the investigation of oxidative/nitrative stress in clinical practice. On the other hand, its high concentration in these fluids may also represent a limitation of using oxidized albumin as a biomarker, because one may wonder whether or not it is an intentional free radical scavenger or if it is simply targeted because of its abundance and high concentration. In this respect, the fact that, surprisingly, nitrated albumin is not a frequent finding in human pathology, despite HSA is the most abundant protein in most biological fluids on a molar basis and contains 18 Tyr residues, may suggest specificity/selectivity and intentionality. Furthermore, the finding that the relative content of 3nitrotyrosine on the albumin molecule was very low when compared with that of other much less concentrated plasma proteins, such as fibrinogen and complement factor D (57), may corroborate this hypothesis.

In conclusion, it is clear from the most recent publications that redox albuminomics is increasingly used to analyze

oxidative albumin modifications occurring in vivo. This approach has a clear applicability to diseases where oxidative stress is involved in their pathophysiology. Occurrence of oxidized albumin in human body fluids easily obtainable without invasive procedures makes oxidized albumin forms potentially useful as diagnostic biomarkers. In addition, because the albumin half-life in plasma is ~ 20 days, increases in its oxidized forms may be indicative of relatively recent oxidative events in human body, and oxidized albumin forms might serve as early biomarkers for human diseases associated to oxidative stress. The challenge in the near future will be to analyze how these oxidative albumin modifications are related to real impact to the body. It will have to be clarified if albumin oxidation can be disease specific or if it just reveals a disease nonspecific increase in oxidative stress as well as if the extent of albumin oxidation can be a possible discriminant to recognize different diseases or, more likely, different stages of a disease. In this sense, the finding that HNA-1 and HNA-2 levels increase in correlation with the degree of renal dysfunction (75, 76), which in turn is paralleled by enhanced oxidative stress, suggests that the redox state of HSA could be a good marker to investigate the progression of, at least some, oxidative stressrelated human diseases. Anyway, this is still an open question.

Acknowledgments

Owing to the strict space limitations, we could not cite all the works relevant to the presented topic. We sincerely apologize to those authors whose work we could not include. Figure 1 was prepared using and combining medical cliparts and illustrations available within the Servier Medical Art section, by courtesy of Servier International.

References

- Agarwal R. On the nature of proteinuria with acute renal injury in patients with chronic kidney disease. Am J Physiol Renal Physiol 288: F265–F271, 2005.
- Agarwal R. Anti-inflammatory effects of short-term pioglitazone therapy in men with advanced diabetic nephropathy. *Am J Physiol Renal Physiol* 290: F600–F605, 2006.
- Aldini G, Vistoli G, Regazzoni L, Gamberoni L, Facino RM, Yamaguchi S, Uchida K, and Carini M. Albumin is the main nucleophilic target of human plasma: a protective role against pro-atherogenic electrophilic reactive carbonyl species? *Chem Res Toxicol* 21: 824–835, 2008.
- Bar-Or D, Heyborne KD, Bar-Or R, Rael LT, Winkler JV, and Navot D. Cysteinylation of maternal plasma albumin and its association with intrauterine growth restriction. *Prenat Diagn* 25: 245–249, 2005.
- 5. Beck JL, Ambahera S, Yong SR, Sheil MM, de Jersey J, and Ralph SF. Direct observation of covalent adducts with Cys34 of human serum albumin using mass spectrometry. *Anal Biochem* 325: 326–336, 2004.
- Boisvert MR, Koski KG, and Skinner CD. Increased oxidative modifications of amniotic fluid albumin in pregnancies associated with gestational diabetes mellitus. *Anal Chem* 82: 1133–1137, 2010.
- Breusing N and Grune T. Biomarkers of protein oxidation from a chemical, biological and medical point of view. Exp Gerontol 45: 733–737, 2010.
- Bruschi M, Petretto A, Candiano G, Musante L, Movilli E, Santucci L, Urbani A, Gusmano R, Verrina E, Cancarini G, Scolari F, and Ghiggeri GM. Determination of the oxido-

redox status of plasma albumin in hemodialysis patients. *J Chromatogr B Analyt Technol Biomed Life Sci* 864: 29–37, 2008.

- Capeillère-Blandin C, Gausson V, Descamps-Latscha B, and Witko-Sarsat V. Biochemical and spectrophotometric significance of advanced oxidized protein products. *Biochim Biophys Acta* 1689: 91–102, 2004.
- Carballal S, Alvarez B, Turell L, Botti H, Freeman BA, and Radi R. Sulfenic acid in human serum albumin. *Amino Acids* 32: 543–551, 2007.
- Colombo G, Aldini G, Orioli M, Colombo R, Giustarini D, Gornati R, Rossi R, Carini M, Milzani A, and Dalle-Donne I. Water-soluble α-β-unsaturated aldehydes of cigarette smoke induce carbonylation in human serum albumin. *Antioxid Redox Signal* 12: 349–364, 2010.
- Curry S, Mandelkov H, Brick P, and Franks N. Crystal structure of human serum albumin complexed with fatty acid reveals an asymmetric distribution of binding sites. *Nat* Struct Biol 5: 827–835, 1998.
- D'Aguanno S, Franciotta D, Lupisella S, Barassi A, Pieragostino D, Lugaresi A, Centonze D, D'Eril GM, Bernardini S, Federici G, and Urbani A. Protein profiling of Guillain-Barrè syndrome cerebrospinal fluid by two-dimensional electrophoresis and mass spectrometry. *Neurosci Lett* 485: 49–54, 2010.
- Dalle-Donne I, Aldini G, Carini M, Colombo R, Rossi R, and Milzani A. Protein carbonylation, cellular dysfunction, and disease progression. J Cell Mol Med 10: 389–406, 2006.
- Dalle-Donne I, Milzani A, Gagliano N, Colombo R, Giustarini D, and Rossi R. Molecular mechanisms and potential clinical significance of S-glutathionylation. *Antioxid Redox Signal* 10: 445–473, 2008.
- Dalle-Donne I, Rossi R, Colombo R, Giustarini D, and Milzani A. Biomarkers of oxidative damage in human disease. Clin Chem 52: 601–623, 2006.
- 17. Dalle-Donne I, Rossi R, Giustarini D, Milzani A, and Colombo R. Protein carbonyl groups as biomarkers of oxidative stress. *Clin Chim Acta* 329: 23–38, 2003.
- Dalle-Donne I, Scaloni A, and Butterfield DA. (Eds). Redox Proteomics: from Protein Modifications to Cellular Dysfunction and Disease. Hoboken, NJ: Wiley, 2006, p. 944.
- Fanali G, di Masi A, Trezza V, Marino M, Fasano M, and Ascenzi P. Human serum albumin: from bench to bedside. Mol Aspects Med 33: 209–290, 2012.
- Fasano M, Curry S, Terreno E, Galliano M, Fanali G, Narciso P, Notari S, and Ascenzi P. The extraordinary ligand binding properties of human serum albumin. *IUBMB Life* 57: 787– 796, 2005.
- Fukushima H, Miwa Y, Shiraki M, Gomi I, Toda K, Kuriyama S, Nakamura H, Wakahara T, Era S, and Moriwaki H.
 Oral branched-chain amino acid supplementation improves the oxidized/reduced albumin ratio in patients with liver cirrhosis. *Hepatol Res* 37: 765–770, 2007.
- 22. Giustarini D, Dalle–Donne I, Lorenzini S, Milzani A, and Rossi R. Age-related influence on thiol, disulfide and protein mixed disulfide levels in human plasma. *J Gerontol A Biol Sci Med Sci* 61: 1030–1038, 2006.
- Giustarini D, Dalle-Donne I, Tsikas D, and Rossi R. Oxidative stress and human diseases: Origin, link, measurement, mechanisms, and biomarkers. Crit Rev Clin Lab Sci 46: 241–281, 2009.
- 24. Greilberger J, Koidl C, Greilberger M, Lamprecht M, Schroecksnadel K, Leblhuber F, Fuchs D, and Oettl K. Malondialdehyde, carbonyl proteins and albumin-disulphide as useful oxidative markers in mild cognitive impairment and Alzheimer's disease. Free Radic Res 42: 633–638, 2008.

- Hackett TL, Scarci M, Zheng L, Tan W, Treasure T, and Warner JA. Oxidative modification of albumin in the parenchymal lung tissue of current smokers with chronic obstructive pulmonary disease. *Respir Res* 11: 180–189, 2010.
- Himmelfarb J and McMonagle E. Albumin is the major plasma protein target of oxidant stress in uremia. *Kidney Int* 60: 358–363, 2001.
- Hortin GL, Seam N, Gerard T, and Hoehn GT. Bound homocysteine, cysteine, and cysteinylglycine distribution between albumin and globulins. Clin Chem 52: 2258–2264, 2006.
- 28. Jiao K, Mandapati S, Skipper PL, Tannenbaum SR, and Wishnok JS. Site-selective nitration of tyrosine in human serum albumin by peroxynitrite. *Anal Biochem* 293: 43–52, 2001.
- Joffe P and Henriksen JH. Bidirectional peritoneal transport of albumin in continuous ambulatory peritoneal dialysis. Nephrol Dial Transplant 10: 1725–1732, 1995.
- Kawai K, Hayashi T, Matsuyama Y, Minami T, and Seiichi E. Difference in redox status of serum and aqueous humor in senile cataract patients as monitored via the albumin thiolredox state. *Jpn J Ophthalmol* 54: 584–588, 2010.
- Kawakami A, Kubota K, Yamada N, Tagami U, Takehana K, Sonaka I, Suzuki E, and Hirayama K. Identification and characterization of oxidized human serum albumin. A slight structural change impairs its ligand-binding and antioxidant functions. FEBS J 273: 3346–3357, 2006.
- 32. Lemarechal H, Allanore Y, Chenevier-Gobeaux C, Kahan A, Ekindjian OG, and Borderie D. Serum protein oxidation in patients with rheumatoid arthritis and effects of infliximab therapy. *Clin Chim Acta* 372: 147–153, 2006.
- Li HY, Hou FF, Zhang X, Chen PY, Liu SX, Feng JX, Liu ZQ, Shan YX, Wang GB, Zhou ZM, Tian JW, and Xie D Advanced oxidation protein products accelerate renal fibrosis in a remnant kidney model. J Am Soc Nephrol 18: 528–538, 2007.
- Lim PS, Cheng YM, and Yang SM. Impairments of the biological properties of serum albumin in patients on haemodialysis. Nephrology 12: 18–24, 2007.
- 35. Liu SX, Hou FF, Guo ZJ, Nagai R, Zhang WR, Liu ZQ, Zhou ZM, Zhou M, Xie D, Wang GB, and Zhang X. Advanced oxidation protein products accelerate atherosclerosis through promoting oxidative stress and inflammation. *Arterioscler Thromb Vasc Biol* 26: 1156–1162, 2006.
- Marsche G, Frank S, Hrzenjak A, Holzer M, Dirnberger S, Wadsack C, Scharnagl H, Stojakovic T, Heinemann A, and Oettl K. Plasma-Advanced Oxidation Protein Products are potent high-density lipoprotein receptor antagonists in vivo. Circ Res 104: 750–757, 2009.
- 37. McCarty DJ. Synovial fluid. In: *Arthritis and Allied Conditions: a Textbook of Rheumatology,* edited by Koopman WJ, 13th edition, vol. 1. Baltimore, MD: Williams & Wilkins, 1997, pp. 81–102.
- Mera K, Anraku M, Kitamura K, Nakajou K, Maruyama T, and Otagiri M. The structure and function of oxidized albumin in hemodialysis patients: Its role in elevated oxidative stress via neutrophil burst. *Biochem Biophys Res Commun* 334: 1322–1328, 2005.
- 39. Mera K, Anraku M, Kitamura K, Nakajou K, Maruyama T, Tomita K, and Otagiri M. Oxidation and carboxy methyl lysine-modification of albumin: possible involvement in the progression of oxidative stress in hemodialysis patients. *Hypertens Res* 28: 973–980, 2005.
- Mitrogianni Z, Barbouti A, Galaris D, and Siamopoulos KC. Oxidative modification of albumin in predialysis, hemodialysis, and peritoneal dialysis patients. Nephron Clin Pract 113: c234–c240, 2009.

41. Moon GJ, Shin DH, Im DS, Bang OY, Nam HS, Lee JH, Joo IS, Huh K, and Gwag BJ. Identification of oxidized serum albumin in the cerebrospinal fluid of ischaemic stroke patients. *Eur J Neurol* 18: 1151–1158, 2011.

- 42. Morgan PE, Sturgess AD, and Davies MJ Evidence for chronically elevated serum protein oxidation in systemic lupus erythematosus patients. *Free Radic Res* 43: 117–127, 2009.
- 43. Murphy MP, Holmgren A, Larsson NG, Halliwell B, Chang CJ, Kalyanaraman B, Rhee SG, Thornalley PJ, Partridge L, Gems D, Nyström T, Belousov V, Schumacker PT, and Winterbourn CC. Unraveling the biological roles of reactive oxygen species. *Cell Metab* 13: 361–366, 2011.
- 44. Musante L, Bruschi M, Candiano G, Petretto A, Dimasi N, Del Boccio P, Urbani A, Rialdi G, and Ghiggeri GM. Characterization of oxidation end product of plasma albumin "in vivo." *Biochem Biophys Res Commun* 349: 668–673, 2006.
- 45. Musante L, Candiano G, Petretto A, Bruschi M, Dimasi N, Caridi G, Pavone B, Del Boccio P, Galliano M, Urbani A, Scolari F, Vincenti F, and Ghiggeri GM. Active focal segmental glomerulosclerosis is associated with massive oxidation of plasma albumin. *J Am Soc Nephrol* 18: 799–810, 2007
- 46. Nagai K, Betsuyaku T, Kondo T, Nasuhara Y, and Nishimura M. Long term smoking with age builds up excessive oxidative stress in bronchoalveolar lavage fluid. *Thorax* 61: 496–502, 2006.
- 47. Nagai K, Betsuyaku T, Konno S, Ito Y, Nasuhara Y, Hizawa N, Kondo T, and Nishimura M. Diversity of protein carbonylation in allergic airway inflammation. *Free Radic Res* 42: 921–929, 2008.
- Noel-Georis I, Bernard A, Falmagne P, and Wattiez R. Database of bronchoalveolar lavage fluid proteins. J Chromatogr B Analyt Technol Biomed Life Sci 771: 221–236, 2002.
- 49. Oettl K and Marsche G. Redox state of human serum albumin in terms of cysteine-34 in health and disease. *Methods Enzymol* 474: 181–195, 2010.
- Oettl K, Stadlbauer V, Krisper P, and Stauber RE. Effect of extracorporeal liver support by molecular adsorbents recirculating system and prometheus on redox state of albumin in acute-on-chronic liver failure. *Ther Apher Dial* 13: 431– 436, 2009.
- 51. Oettl K, Stadlbauer V, Petter F, Greilberger J, Putz-Bankuti C, Hallstrom S, Lackner C, and Stauber RE. Oxidative damage of albumin in advanced liver disease. *Biochim Biophys Acta* 1782: 469–473, 2008.
- 52. Oettl K and Stauber RE. Physiological and pathological changes in the redox state of human serum albumin critically influence its binding properties. *Br J Pharmacol* 151: 580–590, 2007.
- Oettl, K, Reibnegger G, and Schmut O. The redox state of human serum albumin in eye diseases with and without complications. *Acta Ophthalmol* 89: e174–e179, 2011.
- 54. Pavone B, Sirolli V, Bucci S, Libardi F, Felaco P, Amoroso L, Sacchetta P, Urbani A, and Bonomini M. Adsorption and carbonylation of plasma proteins by dialyser membrane material: in vitro and in vivo proteomics investigations. Blood Transfus 3: S113–S119, 2010.
- Pavone B, Sirolli V, Giardinelli A, Bucci S, Forlì F, Di Cesare M, Sacchetta P, Di Pietro N, Pandolfi A, Urbani A, and Bonomini M. Plasma protein carbonylation in chronic uremia. J Nephrol 24: 453–464, 2011.
- Peters T, Jr. The albumin molecule: Structure and chemical properties. In: All About Albumin: Biochemistry, Genetics, and

Medical Applications, edited by Peters T, Jr. San Diego, CA: Academic Press, 1996, pp. 9–75.

- 57. Piroddi M, Palmese A, Pilolli F, Amoresano A, Pucci P, Ronco C, and Galli F. Plasma nitroproteome of kidney disease patients. *Amino Acids* 40: 653–667, 2011.
- 58. Predescu DN, Predescu SA, and Malik AB. Transport of nitrated albumin across continuous vascular endothelium. *Proc Natl Acad Sci U S A* 99: 13932–13937, 2002.
- 59. Quinlan GJ, Mumby S, Martin GS, Bernard GR, Gutteridge JM, and Evans TW. Albumin influences total plasma antioxidant capacity favorably in patients with acute lung injury. Crit Care Med 32: 755–759, 2004.
- 60. Rasheed Z, Ahmad R, and Ali R. Structure and immunological function of oxidised albumin in lung cancer: its potential role as a biomarker of elevated oxidative stress. *Br J Biomed Sci* 66: 67–73, 2009.
- Rasheed Z, Ahmad R, Rasheed N, and Ali R. Reactive oxygen species damaged human serum albumin in patients with hepatocellular carcinoma. *J Exp Clin Cancer Res* 26: 395–404, 2007.
- 62. Rasheed Z and Ali R. Reactive oxygen species damaged human serum albumin in patients with type 1 diabetes mellitus: biochemical and immunological studies. *Life Sci* 79: 2320–2328, 2006.
- 63. Roche M, Rondeau P, Singh NR, Tarnus E, and Bourdon E. The antioxidant properties of serum albumin. *FEBS Lett* 582: 1783–1787, 2008.
- 64. Roche S, Gabelle A, and Lehmann S. Clinical proteomics of the cerebrospinal fluid: towards the discovery of new biomarkers. *Proteomics Clin Appl* 2: 428–436, 2008.
- Rossi R, Giustarini D, Milzani A, and Dalle-Donne I. Cysteinylation and homocysteinylation of plasma protein thiols during ageing of healthy human beings. *J Cell Mol Med* 13: 3131–3140, 2009.
- 66. Selmeci L, Székely M, Soós P, Seres L, Klinga N, Geiger A, and Acsády G. Human blood plasma advanced oxidation protein products (AOPP) correlates with fibrinogen levels. Free Radic Res 40: 952–958, 2006.
- 67. Selmeci L. Advanced oxidation protein products (AOPP): novel uremic toxins, or components of the non-enzymatic antioxidant system of the plasma proteome? *Free Radic Res* 45: 1115–1123, 2011.
- 68. Sheikh Z, Ahmad R, Sheikh N, and Ali R. Enhanced recognition of reactive oxygen species damaged human serum albumin by circulating systemic lupus erythematosus autoantibodies. *Autoimmunity* 40: 512–520, 2007.
- 69. Shi XY, Hou FF, Niu HX, Wang GB, Xie D, Guo ZJ, Zhou ZM, Yang F, Tian JW, and Zhang X. Advanced oxidation protein products promote inflammation in diabetic kidney through activation of renal nicotinamide adenine dinucleotide phosphate oxidase. *Endocrinology* 149: 1829–1839, 2008.
- Soejima A, Matsuzawa N, Hayashi T, Kimura R, Ootsuka T, Fukuoka K, Yamada A, Nagesawa T, and Era S. Alteration of redox state of human serum albumin before and after hemodialysis. *Blood Purif* 22: 525–529, 2004.
- 71. Starosta V and Griese M. Protein oxidation by chronic pulmonary diseases in children. *Pediatr Pulmonol* 41: 67–73, 2006.
- Stewart AJ, Blindauer CA, Berezenko S, Sleep D, Tooth D, and Sadler PJ. Role of Tyr84 in controlling the reactivity of Cys34 of human albumin. FEBS J 272: 353–362, 2005.
- 73. Sugio S, Kashima A, Mochizuki S, Noda M, and Kobayashi K. Crystal structure of human serum albumin at 2.5 A resolution. *Protein Eng* 12: 439–446, 1999.

74. Suzuki M, Betsuyaku T, ItoY, Nagai K, Nasuhara Y, Kaga K, Kondo S, and Nishimura M. Down-regulated NF-E2-related factor 2 in pulmonary macrophages of aged smokers and patients with Chronic Obstructive Pulmonary Disease. *Am J Respir Cell Mol Biol* 39: 673–682, 2008.

- 75. Terawaki H, Takada Y, Era S, Funakoshi Y, Nakayama K, Nakayama M, Ogura M, Ito S, and Hosoya T. The redox state of albumin and serious cardiovascular incidence in hemodialysis patients. *Ther Apher Dial* 14: 465–471, 2010.
- 76. Terawaki H, Yoshimura K, Hasegawa T, Matsuyama Y, Negawa T, Yamada K, Matsushima M, Nakayama M, Hosoya T, and Era S. Oxidative stress is enhanced in correlation with renal dysfunction: examination with the redox state of albumin. *Kidney Int* 66: 1988–1993, 2004.
- 77. Torres MJ, Turell L, Botti H, Antmann L, Carballal S, Ferrer-Sueta G, Radi R, and Alvarez B. Modulation of the reactivity of the thiol of human serum albumin and its sulfenic derivative by fatty acids. *Arch Biochem Biophys* 521: 102–110, 2012.
- 78. Tsikas D, Sandmann J, and Frölich JC. Measurement of S-nitrosoalbumin by gas chromatography-mass spectrometry. III. Quantitative determination in human plasma after specific conversion of the S-nitroso group to nitrite by cysteine and Cu(2+) via intermediate formation of S-nitrosocysteine and nitric oxide. J Chromatogr B Analyt Technol Biomed Life Sci 772: 335–346, 2002.
- Turell L, Botti H, Carballal S, Ferrer-Sueta G, Souza JM, Durán R, Freeman BA, Radi R, and Alvarez B. Reactivity of sulfenic acid in human serum albumin. *Biochemistry* 47: 358– 367, 2008.
- Turell L, Botti H, Carballal S, Radi R, and Alvarez B. Sulfenic acid-A key intermediate in albumin thiol oxidation. J Chromatogr B Analyt Technol Biomed Life Sci 877: 3384–3392, 2009.
- Watanabe A, Matsuzaki S, Moriwaki H, Suzuki K, and Nishiguchi S. Problems in serum albumin measurement and clinical significance of albumin microheterogeneity in cirrhotics. *Nutrition* 20: 351–357, 2004.
- 82. Wayenberg JL, Cavedon C, Ghaddhab C, Lefèvre N, and Bottari SP. Early transient hypoglycemia is associated with increased albumin nitration in the preterm infant. *Neonatology* 100: 387–397, 2011.
- 83. Wayenberg JL, Ransy V, Vermeylen D, Damis E, and Bottari SP. Nitrated plasma albumin as a marker of nitrative stress and neonatal encephalopathy in perinatal asphyxia. *Free Radic Biol Med* 47: 975–982, 2009.
- 84. Zurawa-Janicka D, Renke J, Popadiuk S, Skorko-Glonek J, Szumera M, Plata-Nazar K, Ulko P, Wozniak M, and Lipinska B. Preferential immunoglobulin oxidation in children with juvenile idiopathic arthritis. *Scand J Rheumatol* 35: 193–200, 2006.

Address correspondence to:
 Dr. Isabella Dalle-Donne
 Department of Life Sciences
 University of Milan
 via Celoria 26
 I-20133 Milan
 Italy

E-mail: quack@unimi.it

Date of first submission to ARS Central, May 15, 2012; date of acceptance, May 16, 2012.

Abbreviations Used

2-DE = two-dimensional polyacrylamide gel electrophoresis

AGEs = advanced glycation end products

Alb-CO = carbonylated albumin

Alb-Cys = S-cysteinylated albumin (S-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1])

Alb-SH = mercaptalbumin

Alb-SOH = albumin sulfenic acid

Alb- $SO_2H =$ albumin sulfinic acid (also called human nonmercaptalbumin2 [HNA-2])

Alb-SO₃H = albumin sulfonic acid (also called human nonmercaptalbumin2 [HNA-2])

Alb-SSR = albumin-mixed disulfide (*S*-thiolated albumin, also called human nonmercaptalbumin1 [HNA-1])

AOPPs = advanced oxidation protein products

BALF = bronchoalveolar lavage fluid

COPD = chronic obstructive pulmonary disease

CSF = cerebrospinal fluid

DNP = 2,4-dinitrophenylhydrazone

DNPH = 2,4-dinitrophenylhydrazine

ELISA = enzyme-linked immunosorbent assay

HSA = human serum albumin

LC-ESI-MS/MS = liquid chromatography-electrospray ionization tandem mass spectrometry

MALDI-TOF = matrix-assisted laser desorption/ionization time of flight

RNS = reactive nitrogen species

SDS-PAGE = sodium dodecyl sulfate–polyacrylamide gel electrophoresis

SF = synovial fluid